

Deprotection 2'- ACE Protected RNA

This protocol is for the Dharmacon Deprotection 2'- ACE Protected RNA.

1. Centrifuge tubes briefly.
2. Add 400 µL of 2'- Deprotection buffer to each tube of RNA.
3. Completely dissolve RNA pellet by pipetting up and down.
4. Vortex for 10 seconds and centrifuge for 10 seconds.
5. Incubate at 60 ° C for 30 minutes.
 - Incubate at 60 ° C for 2 hours for oligos with biotin modifications or homopolymer stretches of rA longer than 10 bases.
6. Lyophilize or SpeedVac to dryness before use.
7. The dry pellet can be stored at -20 ° C until use or resuspended in an appropriately buffered RNase-free solution.
8. If appropriate for your application you may perform desalting of the RNA oligo by Ethanol Precipitation. (dharmacon.gelifesciences.com/uploadedfiles/resources/ethanol-precipitation-of-rna-oligonucleotide-protocol.pdf) or desalting columns (dharmacon.gelifesciences.com/uploadedfiles/resources/reverse-phase-protocol.pdf)

Procedure:

2'-Deprotection Buffer is dilute (100 mM) acetic acid adjusted to a pH of 3.4-3.8 using TEMED.

1. Mix 571 µL glacial acetic acid with 99.4 mL RNase-free water to make 100 mL of 100 mM acetic acid.
2. Adjust the pH of the 100 mM acetic acid to 3.4-3.8 using TEMED.

2'-Deprotection buffer = 100 mM acetic acid, adjusted to pH 3.8 with TEMED.

